

Altered glucose and lactic acid contents in the blood and urine of rabbits during aflatoxicosis

A. S. Kolhe

Department of Zoology, Arts and Science College, Bhalod Tal. Yawal Dist. Jalgaon 425304 Maharashtra, India

Article history:

Received 28 July 2025

Received in revised form

30 September 2025

Accepted 02 October 2025

Available online 13 October 2025

Corresponding author Email:
kolheajaykumar4@gmail.com



Abstract

Blood and urine samples were collected on 0,15,30,45,60,75 and 90 days of treatment. Concentration of glucose in both blood and urine were decreased upto 30 days followed with significantly increased value from 45th day onwards. Concentration of lactic acid in blood and urine were significantly increased on 15th day onwards. Increased concentrations of glucose and lactic acid in blood during aflatoxicosis was followed with its increased appearance in urine. It clearly indicates that uptake of glucose and lactic acid by tissues particularly by liver is greatly hampered.

Keywords: Aflatoxin, aflatoxicosis, glucose, lactic acid, blood, urine.

This work is licensed under a Creative Commons Attribution 4.0 International License. This allows re-distribution and re-use of a licensed work on the condition that the author is appropriately credited and the original work is properly cited.

©2025 AASTEMER. All rights reserved.

Introduction

The only nutrition that the brain, retina, and gonad germinal epithelium can use in large enough amounts to provide them with the energy they need is glucose. Thus, one of the body's most intricately con-

trolled homeostatic mechanisms in which the liver plays a significant role is the maintenance of a normal blood glucose level. A "Soskin effect" has been noted in numerous species, wherein the liver absorbs glucose

when the portal glucose concentration climbs above a particular threshold in response to variations in the concentration. and, when it drops below a particular threshold, releasing glucose.

The body retains the large amount of lactic acid produced during anaerobic glycolysis. but once oxygen is available once more. Lactic acid can be utilized directly as fuel or it can be transformed back into glucose. The liver is where the majority of this reconversion takes place.

Aspergillus flavus produces secondary hazardous fungal compounds called aflatoxins. When aflatoxin-contaminated food or feed is consumed, aflatoxicosis, an extremely toxic condition, develops. According to Busby and Wogan,¹ aflatoxin is a strong hepatotoxin that causes hepatocarcinogenesis in a number of species and may be connected to liver cancer in men.^{2,3,4,5} Further more, dietary toxicosis has been linked to biochemical alterations in the liver,⁶ heart,⁷ kidney⁸ and skeletal muscle⁹ in rabbits, resulting in hyperglycemia and a decreased red blood cell count containing PCV.¹⁰ Since concentrations of blood glucose and lactic acid are controlled by interactions of various organs including liver, kidney skeletal muscle. The present investigation was undertaken.

Materials and Methods

Aspergillus parasiticus (NRRL 3240 obtained from I.A.R.I. ,New Delhi, India) was grown on SMKY liquid medium at 280c for 10 days.¹¹ Pooled culture filtrate were extracted with analytical grade chloroform and qualitatively analyzed for different types of aflatoxins on TLC plates¹² aflatoxin content was quantified¹³ using a shimazu UV160A spectrophotometer

The crude aflatoxin concentrate in chloroform was mixed with feed (7.5 mg aflatoxin / kg) and left overnight to allow for chloroform evaporation. The presence of toxin in such toxin mixed ration was ensured by taking random sample and analysis. Feed for control rabbit was prepared in similar way except for addition of toxin.

Young adult New Zealand strain of rabbits (*Oryctolagus cuniculus*) weighing approximately 1.2 kg were maintained under laboratory condition and fed with rations and water ad-libitum. Ten rabbits were randomly segregated in two groups and fed with either control or test feed continuously for 90 days.

Blood (from ear pinna) and urine samples of rabbits were collected for analysis on 0, 15, 30, 45, 60, 75 and 90 days of treatment. The concentrations of glucose¹⁴ and lactic acid¹⁵ were measured in deproteinized samples.

The data were statistically analysed using student's 't' test.

Results

Glucose:

The concentration of glucose in blood of control rabbits was 120.63 mg/dl. Feeding aflatoxin contaminated diet caused initial decline (upto 30 days) followed with a rise. Changes at 15. 30. 45. 75 and 90 days of treatments were found significantly different from control. Concentration of glucose in urine of control rabbits was 93.124 mg/dl. Feeding aflatoxin contaminated diet caused initial decline (upto 30 days) followed with increased appearance of glucose in urine. Changes at 15. 30. 60. 75 and 90 days were found significantly different from control.

Table 1: Effect of feeding aflatoxin contaminated diet on concentration of glucose and lactic acid in blood and urine in rabbits during aflatoxicosis.

Days of treatment	Concentration of glucose (mg/dl)		Concentration of lactic acid (mg/dl)	
	blood	urine	blood	urine
0 (Control)	120.63 ± 7.53	93.12± 6.87	12.27 ± 1.59	6.16 ± 1.26
15	87.71c ± 8.17	65.91d ± 6.72	21.17 ± 0.95	7.03 ± 0.21
30	71.39b ± 8.15	52.61b ± 6.57	25.06a ± 0.45	8.56 ± 0.21
45	91.75c ± 5.05	86.24 ± 6.87	27.31a ± 1.34	11.54b ± 0.39
60	137.21 ± 7.19	148.28a ± 7.02	33.56a ± 2.17	17.54a ± 1.29
75	152.01d ± 7.85	169.50a ± 7.17	39.56a ± 3.54	21.75a ± 1.11
90	162.44b ± 8.69	198.20a ± 7.17	50.64a ± 1 .61	25.31a ± 1.29

Values are mean ± S.E.M.; n = 5.

Significant at : a = P < 0.001 ; b = P <0.01 ; c = P <0.02 ; d = P <0.05

Lactic acid:

Concentrations of lactic acid were 12.27 mg/dl in blood and 6.16 mg/dl in urine of control rabbits. Time dependent increased concentration of lactic acid were observed in aflatoxin fed rabbits. Significantly increased concentrations were recorded in the blood and urine after 30 and 45 days of treatment respectively.

Discussion

Aflatoxin fed diet caused initial decline in concentration of blood glucose upto 30 days. It may be due to decreased feed and water consumption in treated rabbits or due to digestive disturbances,¹⁶ a similar condition to starvation during aflatoxicosis.

But after 30 days of treatment, blood glucose level gradually increased which could be a result of reduced uptake of glucose by tissues, as evidenced by decreased glycogen concentration in liver,⁶ heart⁷ and Kidney.⁸ Smith and Ross¹⁷ also reported depletion of liver glycogen during aflatoxicosis. Time dependent alteration in blood glucose suggests cumulative toxicity during aflatoxicosis. These changes may arise as an effect of aflatoxins on synthetic enzymes or by inhibition of glycogenesis, acceleration of glycogenolysis and depression of glucose transport into hepatocytes. Intraperitoneal administration of toxin to a day-old chick reduce the level of UDP-glucose-glycogen transglucosylase activity.¹⁸

Mechanisms of increased glucose concentration during aflatoxicosis is not clearly understood but it might be due to alteration in hormones or due to stress condition which occurs during aflatoxicosis, and is known to increase blood

sugar by altering insulin action. Hyperglycaemia may be greater in degree and more prolonged in the presence of cirrhosis and some other liver disease.¹⁹ Induced aflatoxicosis in rabbits caused rise in blood glucose level, decrease in glucose uptake by liver and other tissues followed by its appearance in urine. Glucose concentration in the urine of control rabbits was higher; it could be due to voracious feeding nature of rabbits or any other unknown mechanism.

Aflatoxin contaminated diet caused significantly increased concentration of glucose in urine of treated rabbits. It indicates tubular injuries and reduced their reabsorptive capacity. Aflatoxin induced renal injuries, desquamation of epithelial cells of the proximal tubules and necrosis of glomeruli have been reported in kidney during aflatoxicosis.^{20,21,22,23}

Feeding aflatoxin contaminated diet cause increased concentration of lactic acid in blood which

could be due to enhanced rate of anaerobic metabolism in muscle followed by its release into blood stream. Biologically, rabbits during aflatoxicosis became lethargic and showed respiratory troubles.²⁴ Aflatoxin induced significant decrease in RBC count, Hb, PCV,¹⁰ cytotoxicity on RBC in vitro,²⁵ Hyperbilirubinaemia²⁶ and alteration in skeletal muscle.⁹

LDH, which converts lactate to pyruvate, registered an increase in liver during aflatoxicosis.⁶ This suggests an increased capacity of liver to utilize lactate produced intracellularly brought over to the liver through blood. Aflatoxin is well known hepatotoxic agent and altered histoarchitecture. Although LDH activity increased, liver is unable to utilize elevated concentration of circulating lactic acid in the process of Cori cycle causing its accumulation in the blood followed by excretion in the urine.

It is concluded that aflatoxin contaminated diet caused alterations in carbohydrate metabolism during chronic aflatoxicosis in rabbits.

Acknowledgement

The authors are grateful to Prof. Dr. R.J. Verma, Dept. of Zoology, Gujarat University, Ahmedabad for Laboratory Facilities. and also thankful to Dr. M. D. Friesen of the International Agency for Research on Cancer, Lyon, France for providing samples of pure aflatoxins.

Reference

1. Busby, W. F. and Wogan, G. N. Aflatoxins. In: Searle, C.E., Ed., Chemical Carcinogenesis, II, American Chemical Society, Washington DC. 1984, 945-1136.
2. Butler, W. H.; Greenblatt, M. and Lijinsky, W. Carcinogenesis in rats by aflatoxins B1, G1 and B2. *Cancer Res.* 1969, 29(12), 2206-2211.
3. Groopman, J. D.; Cain, L. G. and Kensler, T. W. Aflatoxin exposure in human population: measurements and relationship to cancer. *Crit. Rev. Toxicol.* 1988, 19(2), 113-145.
4. Stoloff, L. Aflatoxin - an overview. In: Rodricks, J. C., Hesseltine, C.W., Mehlman, M.A. (eds.). *Mycotoxins in Human and Animal Health*. Pathotox Publishers, Park Forest South I. 1977, pp. 7-28
5. Kolhe, A. S. and S. R. Ingale. Effect of Mycotoxins in Human and Animal System. *Journal of Research & Development.* 2011, 1(2), 22-28.
6. Kolhe, A. S. Biochemical alterations in liver of

rabbits during aflatoxicosis. *Mukt Shabd Journal.* 2024, 13(1), 820-831.

7. Verma, R. J. and A. S. Kolhe. Effect of dietary aflatoxicosis on biochemical composition of heart. *Proc. Nat. Acad. Sci. India.* 1997, 67(B) III & IV, 239-242.
8. Kolhe, A.S.; R.J. Verma; S.B.Chaudhari and R. V. Bhole. Effects of Dietary Aflatoxicosis of Biochemical composition of kidney. *Research Link.-25.* 2006, IV(8), 134-136.
9. Kolhe, A. S. and Verma, R. J. Aflatoxin in Chewing Product and Their Effect on Skeletal Muscle of Rabbits. *Research Journey.* 2017, 20 Spl.Issue, 74-83.
10. Kolhe, A. S.; S. B.Chaudhari and R. J. Verma. Studies of haematological alterations in rabbits during aflatoxicosis. *Journal of Emerging Technologies and Innovative Research (JETIR).* 2019, 6(2), 308-315.
11. Diener, U. L. and Davis, N. D. Aflatoxin Production by isolates of *Aspergillus flavus*. *Phytopathlogy.* 1966, 56(12), 1390–1393.
12. Reddy, T. V; Vishwanathan, L. and Venkatasubramanian, T. A. Thin layer chromatography of aflatoxins. *Anal Biochem.* 1970, 38(2), 568-571.
13. Nabney, J. and Nesbitt, B. F. A Spectrophotometric Method for Determining the Aflatoxins. *Analyst.* 1965, 90, 155-160.
14. Folin, O. and Malmros, H. An Improved Form of Folin's Micro Method for Blood Sugar Determinations. *J. Biol. Chem.* 1929, 83, 115-120.
15. Barker, S. B. and Summerson, W. H. The Colorimetric Determination of Lactic Acid in Biological Material. *Journal of Biological Chemistry.* 1941, 138, 535-554.
16. Boplet, M. Digestive disorders in chickens submitted to double section of the vagus nerves. *Ann. Pac.Sci. Cameroun.* 1973, 14, 165-176.
17. Smith, J. B. and Ross, H. O. (eds.). *Hycotoxins Formation, Analysis and significance.* John Wiley and Sons. New York. 1985.
18. Shankaran, R.; Raj, H. G. and Venkatasubramanian, T. A. Effect of aflatoxin on carbohydrate metabolism in chick liver. *Enzymologia.* 1970, 39, 371-378.
19. Yeung, R. T. T. and Wang, C. C. L. A study of carbohydrate metabolism in postnecrotic cirrhosis of the liver. *Gut.* 1974, 15, 907-912.
20. Butler, W. H. and Barnes. J.M. Toxic effects of groundnut meal containing aflatoxin to rats and guinea pigs. *Brit. J. Cancer.* 1963, 17, 699-710.
21. Butler, W. H. Acute toxicity of aflatoxin B1 in rats. *Brit.J. Cancer.* 1964, 16, 756-762.
22. Butler, W. H. Acute toxicity of aflatoxin B1 in guinea pigs. *J. Pathol. Bacteriol.* 1966, 91, 277-260.
23. Madhavan, T. V. and Rao, K. S. 1967 Tubular epithelial reflux in the kidney in aflatoxin poisoning. *J. Pathol.Bacterio.* 1967, 93, 329-330.
24. Kolhe, A. S. and Verma, R. J. Effect of Dietary Aflatoxin induced Morphological Alterations in young Rabbits (*Oryctolagus Cuniculus*). *Research Link-38,* 2007, 4(3), 28-29.
25. Verma. R. J. and Raval. P. J. Cytotoxicity of aflatoxin in red blood corpuscles. *Bull. Environ. Contam. Toxicol.* 1991, 47: 426-438.
26. Kolhe, A.S.; R. J. Verma and R. V. Bhole. Effect of Dietary Aflatoxin on Bile Content of Gall Bladder and Bilirubin Concentration During Aflatoxicosis in Rabbits. *Research Link-32,* 2006, 5 (7), 21-22.